


Article

Foliar Calcium Fertilizers Impact on Several Fruit Quality Characteristics and Leaf and Fruit Nutritional Status of the 'Hayward' Kiwifruit Cultivar

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Abstract: Calcium preharvest application influences fruit quality. The impact of preharvest foliar sprays using several commercial fertilizers with Ca content on the fruit quality and nutritional status of the kiwi cv. 'Hayward' was investigated for a 2-year period. Fruit flesh firmness increased under all Ca sprays compared to the control. Total soluble solids and acidity were not altered significantly by any of the studied Ca products. Treatment differences with regard to fruit firmness, soluble solids concentration and acids at harvest were maintained during cold storage for 2 and 4 months. Foliar sprays did not affect the N, P, K, Mg, B, Fe, Mn and Zn concentrations of leaves and fruits. However, all treatments increased the concentration of Ca in leaves and fruits compared to the control.

Keywords: foliar fertilization; fruit firmness; storability

1. Introduction

Kiwifruit is a climacteric fruit with a long postharvest life in cool storage. Calcium (Ca) is a fundamental nutrient related to qualitative characteristics in kiwifruit. Calcium has been referred to as both (i) an essential element and (ii) a nutrient significantly contributing to the maintenance of high postharvest fruit quality.

It also has significant roles in plant physiology, such as a structural function in the cell wall and membranes; a counter ion for inorganic and organic anions in the vacuole; and a cytoplasmic secondary messenger associated with environmental or developmental stimuli and their physiological responses [1].

The Ca effect on the integrity of the cell membrane and its synergetic role with boron (B) in the plant cell wall's build is well documented [2]. The stability of the cell wall relates to the cooperative binding of polygalacturonate chains with Ca ions, forming the cell wall of the fruit interior accessible to enzymes that generate softening or to cell wall denigrative enzymes caused by fungi. Enhancing the fruit Ca content may influence storability by its function in binding the pectin in cell walls and posterior blocking of the access of degrading enzymes, e.g., polygalacturonase [3]. Kiwifruit undergoes physiological disorders during storage [4]. Losses in kiwifruit are mostly due to its relatively high metabolic activity during storage [5]. Fruit softening is frequently ascribed to modifications in cell wall structure

and, specifically, to the effect of pectin de-esterification and Ca crosslink arrangement on cell wall physical characteristics involving strength and elasticity, cell wall loosening and swelling.

The water delivery and cell wall interactions in the apoplast affect, to a great extent, the Ca accumulation and allocation in fruit. Localized Ca deficiencies detected in particular species may originate from alterations in xylem morphology, fruit water relations and pectin composition and can potentially cause porous membranes, irregular cell wall weakening, impaired hormonal signaling and abnormal fruit growth [6].

Calcium is mainly transported by the xylem. High growth together with low transpiration rates in fruits lowers the Ca content, which may fall below the critical level of deficiency, which is necessary for a good cell wall structure and membrane [7]. Consequently, low Ca uptake and transportation via the xylem may cause fruit disorders. In kiwifruit, xylem operating, fruit transpiration, fruit hair viability and fruit hydraulic conductance showed significant alterations during the first 8–10 weeks after full bloom [8]. Each of these factors can be responsible for the primal discontinuance of Ca import into the fruit. Xylem transport happens only in the acropetal direction and is of some significance for the Ca supply of young fruits which yet transpire in the early phase of their development. In later stages, fruits mainly feed from the phloem [9]. The phloem transport system happens through the cytoplasm, which has low concentrations of Ca [10]. Due to its phloem immobility, foliage-applied Ca is not reallocated from treated leaves to the fruit [11].

Early fruit development is the most pivotal period for Ca accumulation in most fleshy fruits, including kiwifruit. Predominantly, this happens due to the fact that Ca is xylem-mobile but phloem-immobile [12]. Thus, the Ca import rate is determined by the xylem sap in the flow rate to the fruit. In the early season, for most of the fleshy fruits, the phloem and xylem saps' influxes are similar [13], while in the late season, the inflow of the total sap is principally dominated by the phloem, with the xylem sap in flow being insignificant due to the transpiration's dynamic decline [14,15]. Hence, in the initial stages of kiwifruit development (approximately from day 10 to day 50 after fruit set), a deduction in atmospheric vapor pressure deficit causes a reduction in several processes such as fruit transpiration, fruit xylem sap inflow and, seemingly, Ca import [16]. Whilst for transpiring organs, their transpiration presumably remains the dominant driver for Ca accretion, within-plant transport and partitioning of Ca and other minerals are also considered to be affected by factors not related to transpiration. For instance, the Ca requirement by an organ and the physicochemical features of the conducting tissues (e.g., ion adsorption and desorption occurring at exchange sites across the walls of the xylem pathway) are probable to impact the metabolic movement of Ca [17]. Furthermore, the action of specialized transfer cells and the higher expression of Ca transporters and Ca binding proteins may advance accumulation of Ca into the fruit unassisted by its transpiration [18,19]. Results indicate that Ca accumulation is coupled to accretive transpiration under high vapor pressure deficit because under that condition, cumulative transpiration equates liken xylem stream. At low vapor pressure deficit, Ca gain by fruit is uncoupled from transpiration because ~60% of the xylemic water is necessary to endure fruit growth [20].

The critical factors affecting Ca delivery and distribution in aerial tissues involve: the rate of xylem water mass flow, the competition between ions for binding sites in xylem vessel walls and pit membranes, formation of lowly soluble or insoluble complexes (e.g., calcium oxalate) and cellular water/ionic transport mechanisms [14,21,22].

Calcium availability in acid soils is low [9]. Although Ca is the most common exchangeable cation in calcareous soils, fruit Ca content is fairly low [23]. Therefore, for specific fruit trees, applying a preharvest foliar Ca spray is a usual practice in order to enhance fruit Ca content. Nevertheless, the impact of foliar Ca spray on the Ca content and the prevalence of disorders in fruit are sometimes discrepant. For efficient foliar Ca application, the physical characteristics of Ca salts can unveil eloquent information. There are three implicit accountings for inconsistencies of Ca treatments: (a) atmospheric effects of Ca

absorption, (b) abnormal Ca distribution of Ca in fruit within the canopy and (c) tree management and cultivation [6].

The aim of this research is to analyze the impact of foliar sprays using several commercial products with Ca content on the fruit quality and nutritional status of the kiwi cv. ‘Hayward’.

2. Materials and Methods

The research was conducted for two consecutive years (2018 and 2019) in a commercial kiwifruit (*Actinidia deliciosa* (A. chev.) C.F. Liang et A.R. Ferguson var. *deliciosa*) orchard located in Naoussa (northern Greece, long. 22°12'0" E; lat. 40°29'04" N; elevation 350 m). The vines of the cultivar ‘Hayward’ (Apostolou Nurseries, Episkopi, Naoussa, Greece) were 13 years old, planted at a spacing of 4 × 3.5 m and trained in a T-bar trellis system [24]. The trees received standard horticultural practices regarding pruning, irrigation and fertilization.

Soil samples from the experimental orchard were collected from a depth of 0–60 cm and analyzed [25]. The soil was characterized as sandy loam, slightly alkaline (pH 7.9), with low electrical conductivity (1.30 mS cm⁻¹), organic matter content 3.5% and calcium carbonate content 13.3%. Concerning nutrients in the soil, their concentrations are shown in Table 1.

Table 1. Soil nutrient concentrations (mg kg⁻¹) of the experimental orchard.

P	K	Ca	Mg	B	Mn	Zn	Fe
39	383	1560	548	0.49	18.41	2.49	13.82

Fertilization of the orchard was performed using soil and leaf analyses data. Five foliar sprays (with 30-day interval) were applied by a hand sprayer (Comfort Backup Sprayer 12 L, Gardena, Ulm, Germany) starting on June 2 with the following products: (1) Profical (% w/v CaO 17, MgO 5, organic matter, 10); (2) Chelan Ca (% w/v N 12, CaO 21, MgO 2.8, Cu 0.014, Fe 0.014, Mn 0.014, Zn 0.014, Mo 0.0014, organic matter 2.8), (3) F-away Ca (% w/w P₂O₅ 20, CaO 5); (4) F-away Ca plus Profical; (5) F-away Ca plus Chelan Ca (Nature S.A. Nea Efessos, Pieria, Greece. Fertilizer doses are shown in Table 2.

Table 2. Fertilizer doses (L t⁻¹).

Fertilizer	Dose
Profical	3
Chelan Ca	2.42
F-away Ca	4
F-away Ca + Profical	3 + 1.8
F-away Ca + Chelan Ca	3 + 1.5

Ca concentration of the spray solution was maintained the same for all treatments. Control vines were sprayed with water. A surfactant was also added (Shinulin: ethoxylated isodecyl alcohol; Farma Chem S.A., Thessaloniki, Greece; 0.3 L per tone).

Leaf samples were collected manually at midsummer (25 July). Each leaf sample consisted of the third leaf past the final fruit on a fruiting lateral. Fruit flesh analysis was carried out on samples taken at harvest. All samples were initially washed once with tap water and twice with distilled water. Leaf samples were dried in a forced draft oven at 68 °C for 72 h and ground in a mill to pass a 30-mesh screen. Nitrogen was determined by Kjeldahl’s procedure, B by the azomethine-H method [26], P by the ammonium phosphovanadomolybdate method [27] and K, Ca, Mg, Fe, Mn and Zn by atomic absorption spectrophotometry. Fruit samples (20 per replication) were collected manually in mid-October (harvest period).

One hundred fruits were collected from the five trees of each replication, and therefore we collected 20 fruit per tree. Fruits were sampled at commercial maturity. Fruits from all treatments were harvested at the same time, based on total soluble solids concentration. Fruits were transported immediately to the laboratory for analyses.

Ten fruits per tree were weighted and evaluated individually for soluble solids (%) after extracting the juice of all fruits. Soluble solids were measured with an Atago PR-1 electronic refractometer (Atago Inc., Bellevue, WA, USA), acidity was measured after titration was measured with 0.1N NaOH [28], flesh firmness was measured with an Effegi penetrometer with an 8-mm tip (Effegi Model FT 327, Alfonsine, Italy) and dry matter was measured according to [29]. Lastly, the remaining 10 fruits per tree were placed into a cooling chamber (+0.5 °C) for four months. For these fruits, the same quality attributes were determined.

The adopted experimental design was a randomized block with five replications of 6 treatments (five vines per replication were used). Differences between means were evaluated by using Tuckey's test ($p \leq 0.05$). Statistical analysis was performed using SPSS statistical package (SPSS Statistics for Windows, Version 17.0. SPSS Inc.; 2008, Chicago, IL, USA).

3. Results

All Ca sprays increased fruit flesh firmness compared to the control (Table 3). In another experiment, kiwifruit cv. 'Hayward', during fruit development, was sprayed up to three times with CaCl₂. Kiwifruits sprayed preharvest with CaCl₂ solutions exhibited lower softening rates compared to the controls at both 20 and 0 °C storage temperatures and quality was maintained [30]. Different formulations of Ca such as CaCl₂ and a commercial product, Calbit, at the application rate of 1% Ca *w/v*, were used by Hashmatt et al. [31]. The treatments were applied five times at weekly intervals at the late fruit developmental stage. Firmness was significantly increased by 22% and the total soluble solids were significantly reduced by 4% due to CaCl₂ compared to the control [32]. The one-time (early, mid or late fruit growing season) spray of CaCl₂ had no significant effect on firmness as compared with the control, whereas the highest fruit firmness was obtained in the four and five foliar sprays treatments [31].

Table 3. Kiwifruit weight, fruit firmness, total soluble solids, total titratable acidity and dry matter during harvest (means of 2 years).

Treatments	Fruit Firmness (kg cm ⁻²)	Total Soluble Solids (%)	Total Titratable Acidity (% Citric Acid)	Mean Fruit wt (g)	Dry Matter (%)
Control	6.49 c ^z	6.86 a	1.24 a	128 a	17.15 b
Profical	7.44 ab	6.86 a	1.25 a	124 a	17.35 b
Chelan Ca	7.44 ab	6.79 a	1.33 a	126 a	16.96 b
F-away Ca	7.52 a	6.90 a	1.27 a	124 a	16.88 b
F-away Ca + Profical	7.58 a	6.92 a	1.28 a	126 a	18.67 a
F-away Ca + Chelan Ca	7.57 a	6.82 a	1.24 a	125 a	19.02 a

^z Means within columns and years followed by common letters are not significantly different (Tukey's test, $p \leq 0.05$).

The mean fruit weight was not affected by the foliar application of all Ca products compared to control. Koutinas et al (2010) observed that preharvest foliar sprays with various commercial products containing Ca, over a 2- year period, affected significantly various fruit quality attributes and nutritional status of 'Tshechlidis' kiwifruit cultivar [33]. Foliar application of all Ca products tested plus boron did not affect mean fruit weight compared with the control.

None of the studied Ca products significantly altered total soluble solids and acidity. The total soluble solids were reduced by 4% due to CaCl₂ compared to the control [31]. Dry matter content of fruits was improved after application of F-away Ca + Profical and

F-away Ca + Chelan Ca compared to the rest of the treatments. Fruit dry matter content was non-significantly increased by 3 and 7% due to CaCl_2 and Calbit respectively, compared to the control [31].

We further examined fruit quality attributes of kiwis stored for 2 and 4 months in cooling chambers. The fruit soluble solids concentration increased, whereas total titratable acidity decreased with respect to the harvest stage (Tables 4 and 5). The decay percentage of kiwifruit significantly suppressed by thrice CaCl_2 sprays [32]. Furthermore, fruit firmness slightly decreased during 90 days of cold storage from 7.89 to 4.99 kg cm^{-2} according to the previous authors.

Table 4. Fruit firmness, total soluble solids and total titratable acidity after two months of cold storage (means of 2 years).

Treatments	Fruit Firmness (kg cm^{-2})	Total Soluble Solids (%)	Total Titratable Acidity (% Citric Acid)
Control	4.07 b ^z	12.06 a	1.00 a
Profical	5.05 a	11.93 a	1.07 a
Chelan Ca	5.23 a	12.22 a	1.05 a
F-away Ca	5.22 a	12.29 a	1.03 a
F-away Ca + Profical	5.28 a	11.75 a	1.04 a
F-away Ca + Chelan Ca	5.32 a	12.05 a	1.04 a

^z Means within columns and years followed by common letters are not significantly different (Tukey's test, $p \leq 0.05$).

Table 5. Fruit firmness, total soluble solids and total titratable acidity after four months of cold storage (means of 2 years).

Treatments	Fruit Firmness (kg cm^{-2})	Total Soluble Solids (%)	Total Titratable Acidity (% Citric Acid)
Control	2.16 b ^z	14.10 a	0.82 a
Profical	3.52 a	14.06 a	0.80 a
Chelan Ca	3.73 a	14.18 a	0.81 a
F-away Ca	3.74 a	14.31 a	0.77 a
F-away Ca + Profical	3.92 a	14.10 a	0.84 a
F-away Ca + Chelan Ca	3.91 a	14.42 a	0.83 a

^z Means within columns and years followed by common letters are not significantly different (Tukey's test, $p \leq 0.05$).

Regarding the results of various treatments on fruit firmness, soluble solids concentration and acids at harvest were maintained during cold storage for 2 and 4 months. Fruit softening and water loss are the most important limiting factors during cold storage [33]. During storage, kiwifruit softens noticeably from 6 to 8 kg cm^{-2} at harvest to less than 1 when eaten ripe. The duration of the softening process determines the commercial life of fruit. Various preharvest factors influence the kiwifruit firmness as well as fruit-handling procedures [34]. Increasing fruit Ca concentration through preharvest foliar and fruit sprays with Ca solutions is one of the methods used to preserve firmness and quality characteristics during ripening and storage of fresh fruits [35,36].

Foliar sprays did not affect the N, P, K, Mg, B, Fe, Mn and Zn concentrations of leaves and fruits (Tables 6 and 7). However, all treatments increased the concentration of Ca in leaves and fruits compared to the control. In vines sprayed 17 times with CaCl_2 at 0.8% of the commercial product (1700 ppm Ca), fruit Ca content was higher and storage life was 50–80% greater than fruit from vines sprayed nine times and in fruit from control vines [37]. CaCl_2 sprays increased fruit pericarp, core and skin Ca as reported by Gerasopoulos et al. (1996), who found that fruit pericarp, core and skin Ca was increased by CaCl_2 sprays [30]. Fruit Ca content and other fruit quality characteristics were improved by foliar Ca application [30]. Foliar Ca application was beneficial to increase fruit Ca content and other fruit quality characteristics [31].

Table 6. Effect of foliar sprays on nitrogen (N), phosphorus (P), potassium (K), calcium (Ca), magnesium (Mg), boron (B), manganese (Mn), zinc (Zn) and iron (Fe) concentrations of the kiwifruit fruits (means of 2 years).

Treatments	N (mg g ⁻¹ dry wt)	P (mg g ⁻¹ dry wt)	K (mg g ⁻¹ dry wt)	Ca (mg g ⁻¹ dry wt)	Mg (mg g ⁻¹ dry wt)	B (µg g ⁻¹ dry wt)	Mn (µg g ⁻¹ dry wt)	Zn (µg g ⁻¹ dry wt)	Fe (µg g ⁻¹ dry wt)
Control	1.08 a ^z	0.59 a	1.58 a	0.20 b	0.12 a	16 a	3 a	5 a	13 a
Profical	1.13 a	0.57 a	1.64 a	0.26 a	0.12 a	16 a	4 a	7 a	13 a
Chelan Ca	1.06 a	0.57 a	1.61 a	0.27 a	0.12 a	18 a	4 a	6 a	12 a
F-away Ca	1.08 a	0.58 a	1.75 a	0.27 a	0.13 a	17 a	4 a	7 a	13 a
F-away Ca + Profical	1.12 a	0.60 a	1.69 a	0.28 a	0.13 a	18 a	4 a	6 a	13 a
F-away Ca + Chelan Ca	1.04 a	0.58 a	1.68 a	0.28 a	0.12 a	16 a	4 a	6 a	12 a

^z Means within columns and years followed by common letters are not significantly different (Tukey's test, $p \leq 0.05$).

Table 7. Effect of foliar sprays on nitrogen (N), phosphorus (P), potassium (K), calcium (Ca), magnesium (Mg), boron (B), manganese (Mn), zinc (Zn) and iron (Fe) concentrations of the kiwifruit leaves (means of 2 years).

Treatments	N (mg g ⁻¹ dry wt)	P (mg g ⁻¹ dry wt)	K (mg g ⁻¹ dry wt)	Ca (mg g ⁻¹ dry wt)	Mg (mg g ⁻¹ dry wt)	B (µg g ⁻¹ dry wt)	Mn (µg g ⁻¹ dry wt)	Zn (µg g ⁻¹ dry wt)	Fe (µg g ⁻¹ dry wt)
Control	2.06 a ^z	0.55 a	2.99 a	4.25 b	0.78 a	61 a	60 a	23 a	110 a
Profical	2.15 a	0.59 a	2.93 a	4.72 a	0.75 a	57 a	62 a	24 a	105 a
Chelan Ca	1.97 a	0.55 a	2.96 a	4.8 a ²	0.79 a	61 a	57 a	25 a	108 a
F-away Ca	2.01 a	0.59 a	2.79 a	4.92 a	0.80 a	61 a	64 a	25 a	114 a
F-away Ca + Profical	2.17 a	0.60 a	2.87 a	4.88 a	0.77 a	55 a	59 a	28 a	111 a
F-away Ca + chelan Ca	2.16 a	0.64 a	2.86 a	5.01 a	0.79 a	59 a	58 a	24 a	103a

^z Means within columns and years followed by common letters are not significantly different (Tukey's test, $p \leq 0.05$).

In the kiwifruit cultivar 'Tschelidis', it was found that foliar sprays with fertilizers containing Ca increased the Ca concentration of leaves compared to the control [33]. The increase in the Ca concentration of fruits compared to the control was found after the foliar application of Calfruit plus Qualityfruit, Chelan CaP, Power Ca and Acid CaLMg.

Schonherr (2001) [38] studied cuticular penetration using Ca salts. Humidity over cuticles and hygroscopicity of salts significantly affected the rates of penetration. A prerequisite for penetration of ions is the dissolution of the salt, which was determined by the point of deliquescence (POD) of the salt and humidity over the salt residue. Schonherr (2001) [38] concluded that most suitable salts for foliar nutrition should have a low POD such as CaCl₂ (33%) and Ca(NO₃)₂ (56%), whereas salts with a POD above 90% (for example, Ca-acetate, Ca-lactate and Ca-propionate) are not suitable for foliar nutrition, since penetration causes a humidity rate close to 100%.

Foliar absorption is affected by the presence of N and amino acids in fertilizers since it is reported that urea promotes the simultaneous influx of other nutrients at both the cuticular and cellular levels [39]. Phloem transport of N occurs mainly in the form of amino acids. Furuya and Umemiya (2002) [40] studied the absorption of N into peach leaves through 18 kinds of foliar-applied N chemical forms for the absorption of N into peach leaves. They reported that urea and inorganic N sources, especially nitrate-N, were superior as compared to amino acids with respect to N absorption into the peach leaves. It seems that the composition and the form of Ca (organic, inorganic, etc.) of these forms of N exert a significant role in Ca absorption and transportation.

4. Conclusions

In conclusion, the application of the studied commercial Ca fertilizers was beneficial for kiwifruit, since it improved fruit firmness and increased the Ca concentration of leaves and fruits, as well as fruit dry matter at "F-away Ca + Profical" and "F-away Ca + Chelan" Ca treatments. Treatment differences with regard to fruit firmness, soluble solids concentration and acids at harvest were maintained during cold storage for 2 and 4 months.

Total soluble solids, total titratable acidity and the mean fruit weight were not affected by the foliar sprays of Ca-containing commercial fertilizers. Finally, these products could be applied as foliar sprays during the growing season starting from fruit set up to prior to harvest as they improve the fruit firmness and storability of kiwifruit.

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