

Review

Is the Artificial Pollination of Walnut Trees with Drones Able to Minimize the Presence of *Xanthomonas arboricola* pv. *juglandis*? A Review

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Abstract: Walnut (*Juglans regia* L.) is a monoecious species and although it exhibits self-compatibility, it presents incomplete overlap of pollen shed and female receptivity. Thus, cross-pollination is prerequisite for optimal fruit production. Cross-pollination can occur naturally by wind, insects, artificially, or by hand. Pollen has been recognized as one possible pathway for *Xanthomonas arboricola* pv. *juglandis* infection, a pathogenic bacterium responsible for walnut blight disease. Other than the well-known cultural and chemical control practices, artificial pollination technologies with the use of drones could be a successful tool for walnut blight disease management in orchards. Drones may carry pollen and release it over crops or mimic the actions of bees and other pollinators. Although this new pollination technology could be regarded as a promising tool, pollen germination and knowledge of pollen as a potential pathway for the dissemination of bacterial diseases remain crucial information for the development and production of aerial pollinator robots for walnut trees. Thus, our purpose was to describe a pollination model with fundamental components, including the identification of the “core” pollen microbiota, the use of drones for artificial pollination as a successful tool for managing walnut blight disease, specifying an appropriate flower pollination algorithm, design of an autonomous precision pollination robot, and minimizing the average errors of flower pollination algorithm parameters through machine learning and meta-heuristic algorithms.

Keywords: cross-pollination; *Juglans regia*; literature review; self-compatibility; walnut blight disease; aerial pollination; artificial pollination technologies; pollination drone



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1. Introduction

The process of pollination involves the transfer of pollen grains from the anther, the male portion of a flower, to the female part (stigma) of the same or another plant. There are two types of pollination: (i) self-pollination (autogamy), in which pollen is deposited on the stigma of the same or another flower on the same plant; and (ii) cross-pollination (alogamy), in which pollen is transferred from one plant to the flower of a genetically different plant or cultivar. Walnut trees are self-compatible, but they require cross-pollination with another walnut tree to produce nuts due to being protandrous, meaning that the male flowers mature—release pollen—before the female flowers become receptive, and pollen shedding occurs before female bloom begins, or being protogynous, meaning that the female flower begins opening—become receptive—prior to pollen shedding. Cross-pollination involves transferring pollen from the male flowers of one walnut tree to the female flowers of another. This can occur naturally, with the help of the wind (wind-blown pollen) or insects such as bees, or artificially through the process of hand pollination [1].

On walnut trees, the pollen is produced in catkins, which may also be colonized with *Xanthomonas arboricola* pv. *juglandis*, the causative agent of walnut blight disease [2,3]. Mainly the disease can affect the leaves, stems, and nuts of the tree, leading to reduced yield and poor-quality nuts. *Xanthomonas arboricola* pv. *juglandis* is primarily spread by water, either through rain or irrigation, and can be more severe during warm and wet weather [4,5].

Once the bacterium enters the tree tissues, it can survive and spread within the tree through the sap. To prevent and control *X. arboricola* pv. *juglandis*, farmers can use a combination of cultural and chemical control measures. This may include removing infected plant material, managing irrigation practices to reduce water on the foliage, and applying copper-based fungicides during the dormant season or at the first signs of infection. Here, it is important to note that the overuse of chemical control measures can lead to the development of resistant strains of the bacterium, so it is important to use a holistic approach to disease management that incorporates a variety of methods. Additionally, early detection and rapid response to infected trees can help to limit the spread of the disease to other trees in the orchard.

Research has shown that inoculum of *X. arboricola* pv. *juglandis* is also disseminated through pollen [3,6]. This evidence-based early detection of developmental-behavioral problems from primary infection of *X. arboricola* pv. *juglandis* in symptomless plant materials, such as catkins and pollen, should be considered besides that in symptomatic materials (leaves, fruit, and other plant debris).

Pathogenic bacteria and pollen are two distinct biological entities that are not directly related to each other. While pathogenic bacteria and pollen are not directly related, they can sometimes interact in certain situations. For example, pollen can sometimes serve as a carrier for pathogenic bacteria. Pollen of *Actinidia* spp. is a good substrate for *Pseudomonas syringae* pv. *actinidiae* colonization, survival, and a pathway for its spread [7]. Therefore, pollen may be a conduit for the spread of *P. syringae* pv. *actinidiae* and kiwifruit bacterial canker [7]. Further, pollen dispersal has been presented for *Erwinia amylovora* of pome fruit [8] and for *X. arboricola* pv. *juglandis* of walnut [9,10]. Moreover, pollen was recognized as a possible pathway for *X. arboricola* pv. *juglandis*, although further studies were suggested to confirm if pollen could spread the pathogen [3]. Viable pathogenic bacteria such as *X. arboricola* pv. *juglandis* have often been isolated from pollen, and infected pollen also represents a potential route of introduction into healthy walnut groves even in case of mechanical pollination [3,6]. Walnut cv. Chandler and cv. Serr results showed that pollen was recognized as a possible pathway for *X. arboricola* pv. *juglandis* [3].

Therefore, the survival of the bacterium *X. arboricola* pv. *juglandis* in catkin buds and catkins should be checked before the bacterium overwinters in blighted leaf and catkin buds. When water carries the bacteria to the leaves and stigmatic surfaces of the flowers, blight spreads from these reservoirs. When catkins open and staminate flowers release pollen, the disease may also spread through the air. Contaminated pollen may be carried by wind and deposited on any young growing parts of trees [4].

Artificial pollination technologies have been developed to address various challenges in natural pollination, including the issue of contaminated pollen or contributing to possible disease outbreaks from contaminated pollen [11,12]. Artificial pollination systems have been developed to supplement natural pollination and provide yield security even in poor pollination scenarios. These technologies include various methods for pollen collection and application, such as hand pollination, handheld and vehicle-mounted devices, unmanned aerial vehicles (UAVs), and robotic and autonomous pollinators. These systems have been found to perform adequately for certain crops, including kiwifruit, olive, date palm, walnut, and tomato [13,14].

Moreover, artificial pollination technologies have seen significant advancements, with the integration of machine learning and artificial intelligence (AI) playing a crucial role in the development of innovative solutions for future farming. Especially, AI and machine learning are being integrated into pollination technologies to develop autonomous pollina-

tion systems [15,16]. Based on the above, researchers are developing various drone-based pollination technologies, such as the use of tiny drones equipped with artificial intelligence to autonomously navigate and carry pollen between plants [17,18].

In our view, the parameters are important and will be further analyzed in this review paper. Further, while pollen-mediated transmission of bacterial pathogens has been suggested for *Xanthomonas arboricola* pv. *juglandis*, we will try to answer what are the advantages of pollination drones and how they can help to prevent or reduce the risk of walnut blight disease. In the end, we have designed a path-planning algorithm for a pollination robot that involves determining the disease inoculum and an optimal route for the robot to pollinate flowers efficiently.

2. Walnut Blight Prevention

Walnut Blight and Conditions

Typically, the cycles of walnut blight bacteria are dependent upon weather conditions and rainfall during the growing season. Infrequent rainfall during the spring may lead to monocyclic progress, while frequent spring rainfall tends to favor polycyclic disease epidemics. Rainfalls during late spring (after leaf growth) have been reported to favor the spread of *X. arboricola* pv. *juglandis* bacterium, which causes serious damage to trees and is responsible for significant crop losses, which can reach more than 50% of nut drop [19].

All aerial walnut organs, including catkins, female flowers, leaves, and fruit, are infected [3]. Necrotic lesions on the fruit, twigs, and foliage are characteristic disease symptoms. Leaf lesions consist of small water-soaked spots, surrounded by chlorotic halos that extend to become brown necrotic lesions. Fruit lesions begin as tiny, water-soaked spots and develop into pericarp and inner nut tissue necrosis, which results in early fruit drop. After shell hardening, infections often only impact the epicarp [20].

Cankers serve as a source of inoculum for leaves and nutlet infections [4,21,22]. Populations of *X. arboricola* pv. *juglandis* found on dormant buds serve as the primary inoculum for nut infections [21]. Tissues that have recently been infected can act as secondary sources of inoculum for the pathogen. Pollen released from infected catkins plays a role in pathogen dissemination [2,21]. The bacterium is transmitted through moisture, particularly through the combined action of wind and rain [23]. The cultivars Chandler and Vina exhibit significant susceptibility to *X. arboricola* pv. *juglandis*, thus demonstrating the potential of these cultivars to be infected with *X. arboricola* pv. *juglandis* under conditions favorable for the disease [5,21,24]. The terminal fruitfulness cultivars Milotai, Marbot, Sibisel, and Ronde de Montignac and the lateral fruitfulness cultivars Chandler, Sunland, and Techama were found to be highly susceptible cultivars, making it possible to serve as host responses to bacterial blight infections at different leaf and fruit growth stages [20].

Overall, the degree of infection caused by *X. arboricola* pv. *juglandis* bacterium depends on: (a) the quantity of the pathogen present in individual catkin buds and catkins (inoculum); (b) the quantity of walnut blight cankers present on certain walnut varieties; (c) the environmental conditions, such as rain, which play a significant role in spreading bacteria and aiding infection; and (d) the variety, with early leafing varieties being most severely affected.

Besides the favorable conditions for the disease, research suggests that pollen released from infected catkins plays a role in pathogen dissemination [21,22]. Aerial dissemination of infected pollen from diseased catkins may also transmit the bacterium *X. arboricola* pv. *juglandis* to pistillate flowers. However, this source of inoculum might be region specific [25] or due to different origins of the propagation material [3]. Up to date new evidence provides data that infections depend on pollen, especially in walnut orchards with varieties for which the catkins emerge before the pistillate flowers, i.e., cv. Chandler. Indeed, pollen is important for spreading bacteria and aiding infection [3]. Even more, the isolation of *X. arboricola* pv. *juglandis* in late winter-early spring led to the finding that the primary inoculum is present in buds (overwintering), catkins, and female flowers [4,21].

So, it is crucial to select walnut varieties resistant to bacterial blight and to implement good orchard management techniques, such as pruning and fertilization, to keep trees healthy and less susceptible to disease. Moreover, one should be aware of pollen released from infected catkins, which contributes to the spread of the pathogen, *X. arboricola* pv. *juglandis*. If bacterial blight is present in the orchard, it may be necessary to apply fungicides or use other treatments to prevent the disease from spreading, such as artificial techniques for cross-pollination, which probably directly causes or prevents bacterial blight in walnut trees. Apart from the infection of catkins with *X. arboricola* pv. *juglandis*, which plays a role in pathogen dissemination, walnuts produce pollen that is desiccation intolerant. Pollen that does not have homeostatic mechanisms for maintaining a constant water content dies rapidly after opening of the anther or after pollen dispersal [26]. The previously described ‘partially hydrated’ or, more precisely, desiccation-sensitive pollen of this type may serve as the connection point with *X. arboricola* pv. *juglandis* when the environmental conditions such as temperature and relative humidity affect pollination processes or are highly favorable for pathogen growth in plants [26].

However, the relationship between this pollen property and walnut blight disease during the flower cycle when the daily temperature and leaf wetness are more favorable for *X. arboricola* pv. *juglandis* has not been supported with detailed experimental data. The epiphytic colonization of the stigmas of the kiwifruit flower after inoculation by pollen contaminated with GFPuv-labeled *Pseudomonas syringae* pv. *actinidae* (Psa), which is responsible for the bacterial canker of the kiwifruit, indicated that Psa is often transmitted to the stigma by pollen contamination [27]. Further, it is well known that plants employ sexual mimicry, and flowers mimic the mating signals of their pollinator insects. Based on the mimicry phenomenon, researchers demonstrated that fire blight, a serious disease of pear and apple trees, requires the combination of warm temperatures, open blossoms, and wet weather. The disease spreads quickly from flower to flower through wind, water, and pollinating insects. So, the specific mimic odors that serve as a tool to enhance honeybee foraging and pollination activities in pear and apple crops spread serious diseases such as a fire blight of pear and apple trees [28,29]. So, concerning walnut blight disease, the fundamental questions are: Is flower/corolla closure linked to a decrease in viability of desiccation-sensitive pollen? Is this disease related to other pollination activities or phenomena such as mimicry or dissemination by wind?

As far as walnut plants are concerned, if pollen serves as a possible pathway for the dissemination of *X. arboricola* pv. *juglandis* and walnut blight disease, to prevent the disease from spreading, apart from fungicides, it is necessary to use artificial techniques for cross-pollination, as mentioned above, with uninfected pollen (pollen that is free from the above pathogens or diseases).

3. Walnut Buds: Bloom and Pollination Events—Cross-Pollination

Walnuts are monoecious and have male and female flowers on the same tree. Male flowers are formed in structures called catkins. They develop directly on the prior year’s growth and are easy to identify by eye. At leaf out, the preformed shoot develops, and compound leaves start to grow before the pistillate (female) flowers form. This explains why walnut cultivars are indicated by both their leaf out date and bloom date. Female flowers are only receptive to pollination for a short time. In particular, pollen shed from staminate (male) flowers during anthesis is spread by wind (wind-blown pollen) and remains viable for a short time, generally up to 48 h. As already mentioned, walnut is self-compatible but has adapted a mechanism called dichogamy to reduce the degree of inbreeding. Walnuts are classified as heterodichogamous; however, the majority of commercial walnut cultivars, such as Chandler and Serr, are protandrous, meaning that the male flowers mature and pollen shed occurs before the female bloom begins. So, protandrous cultivars, i.e., Serr, may be large contributors to protogynous cultivar pollination. In most walnut-growing regions, dichogamy is the reason for including pollinizers in orchards. Moreover, in certain

cultivars, such as Serr, excessive pollen on pistillate flowers (too much pollen) has been identified to be the cause of pistillate flower abortion [1].

During cross-pollination, pollen is transferred from the male reproductive organ (stamen) of one plant to the female reproductive organ (pistil) of another plant of the same species, resulting in fertilization and the production of seeds. In the case of nut trees like walnuts, cross-pollination is necessary for the trees to produce nuts. Cross-pollination can occur naturally, through the action of wind or insects such as bees, or it can be performed artificially through hand pollination, in which pollen is manually transferred from one tree to another. Natural cross-pollination can be affected by factors such as the proximity of the trees, the timing of flowering, and the presence of pollinators.

Walnuts require cross-pollination for optimal nut production. Cross-pollination increases the genetic diversity of the trees and can lead to larger and more flavorful fruit. However, not all nut tree varieties are compatible for cross-pollination. It is important to choose compatible varieties and to plant them close enough for natural cross-pollination to occur. In some cases, artificial pollination may be necessary to ensure adequate pollination and fruit set. This is especially true in orchards where natural pollinators are scarce or when the weather is unfavorable for pollination. Artificial pollination involves manually transferring pollen from the male flowers of one tree to the female flowers of another. As mentioned above, walnut trees probably require cross-pollination with another walnut tree to produce nuts and minimize the risk of bacterial contamination. This can occur naturally, artificially with air vehicles, or through the process of hand pollination or the design of specific aerial pollination systems for walnut trees.

4. Artificial Pollination—Artificial Pollination Technology

Pollination is critical for many crops for successful production. To avoid pollination failure due to weather events, bloom asynchrony, insects, and wind-based pollination systems, artificial pollination systems are required that can provide yield security for crops [30].

There are various reasons for resorting to artificial pollination, including:

1. Crop yield enhancement: In agriculture, artificial pollination is sometimes employed to increase crop yields. This can be particularly important for crops where natural pollination may be insufficient [31].
2. Control of pollination: Artificial pollination allows for precise control over the pollination process. This is useful in hybrid seed production, where specific traits can be selected [32].
3. Overcoming pollination challenges: Some crops may face challenges with natural pollination due to factors like low insect activity, unfavorable weather conditions, i.e., wind, or geographic isolation. Artificial pollination can overcome these challenges [33].
4. Biotechnological research: In scientific research, artificial pollination can be used to study plant genetics, breeding, and other aspects of plant biology [34].

Artificial pollination technology involves various methods and tools designed to facilitate the pollination process in plants by assisting the natural transfer of pollen. Some of the technologies and techniques used for artificial pollination include:

1. Drones: Unmanned aerial vehicles (UAVs), or drones, equipped with special devices can be used for pollination. These devices may carry pollen and release it over crops, i.e., walnut trees, or mimic the actions of bees and other pollinators [35].
2. Robotic pollinators: Small robots designed to mimic the behaviors of natural pollinators can navigate through crops, transferring pollen between flowers. These robots are often equipped with cameras and sensors to identify and locate flowers [36].
3. Spraying devices: Some systems involve the use of sprayers to disperse pollen over crops. These devices can be mounted on tractors or other vehicles, releasing pollen in a controlled manner [37].

tion of bacterial diseases of other crops, such as pear and kiwifruit plants (co-keywords: “pear” and “*Actinidia deliciosa*”, Figure 2), remain crucial information for the design and manufacture of aerial pollinator robots for walnut trees.

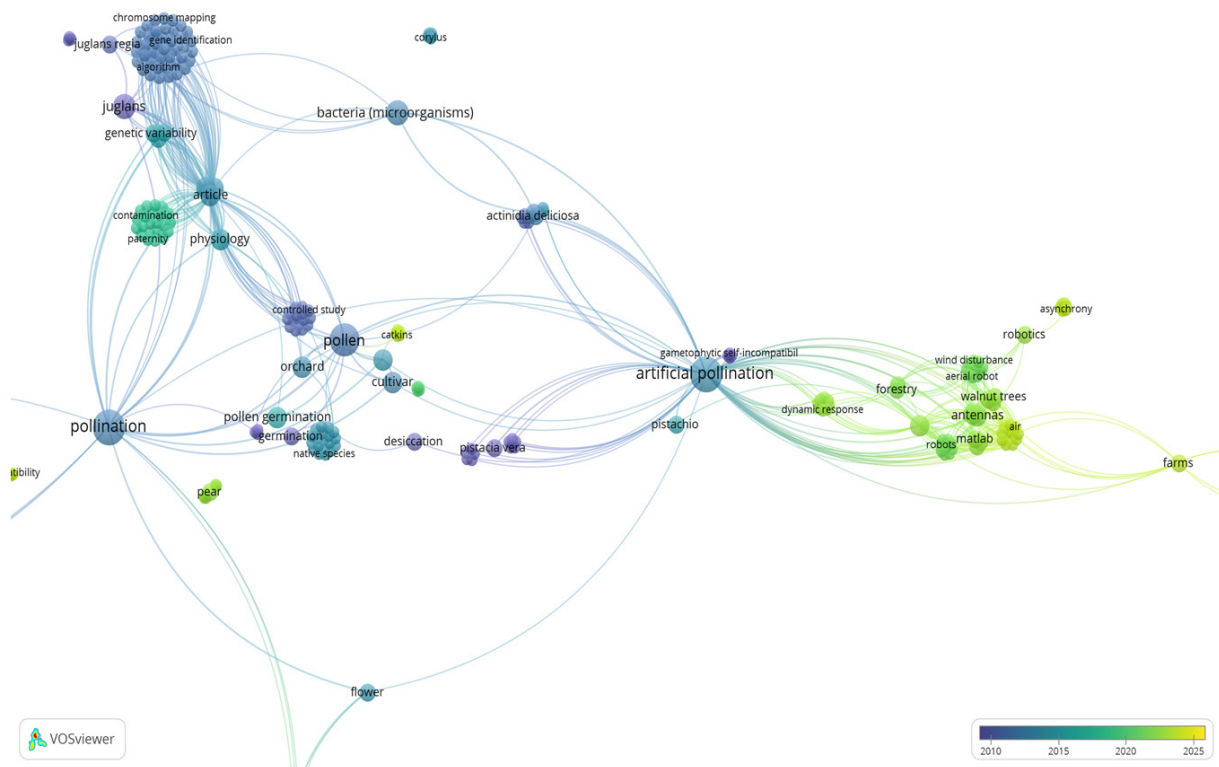


Figure 2. Co-keyword network visualization based on “Artificial” and “Pollination and Technologies and Walnut”.

The keywords “Artificial” and “Pollination and Technologies and Walnut” are presented as 13 clusters defined by 310 keywords (items), which contribute to a total 100%, as presented in Figure 2.

Cluster 1 (Figure 3) is defined by 52 keywords (items), with keywords including “MATLAB”, “Adams MATLAB cosimulation”, “computational fluid dynamics”, “cosimulation”, “flight simulators”, “flying robots”, “quad rotors”, “aerial robot”, “robotics”, “agricultural robots”, “system stability”, “vibration analysis”, “vibration transmissibility”, “software testing”, “antennas”, “artificial pollinations”, “fruit production”, “population growth”, “population statistics”, “walnut pollinator”, “crop growth”, “wind disturbance”, etc.

Cluster 2 (Figure 4) is defined by 45 keywords (items), with keywords including “algorithms”, “bacterial artificial chromosome”, “chromosomes, artificial, bacterial”, “contig mapping”, “expressed sequence tags”, “genome analysis”, “genotyping techniques”, “heterozygosity”, “homozygosity”, “physical map”, “plant genome”, “plant growth”, “pollen sources”, “population genetic structure”, “sequence analysis, DNA”, “single nucleotide polymorphism”, “vegetative propagation”, etc.

Based on the above observations in clusters 1 and 2 (Figures 3 and 4), “artificial pollination technologies” include knowledge of: (a) mechanical pollination technology, i.e., aerial vehicles (UAVs), robotics, and autonomous pollinators (hardware) and basic simulation methodologies for controlling a quadrotor (software); and (b) Mother Nature’s pollination technology, i.e., pollen (pollen germination), crops (cultivars), and microorganisms (bacteria).

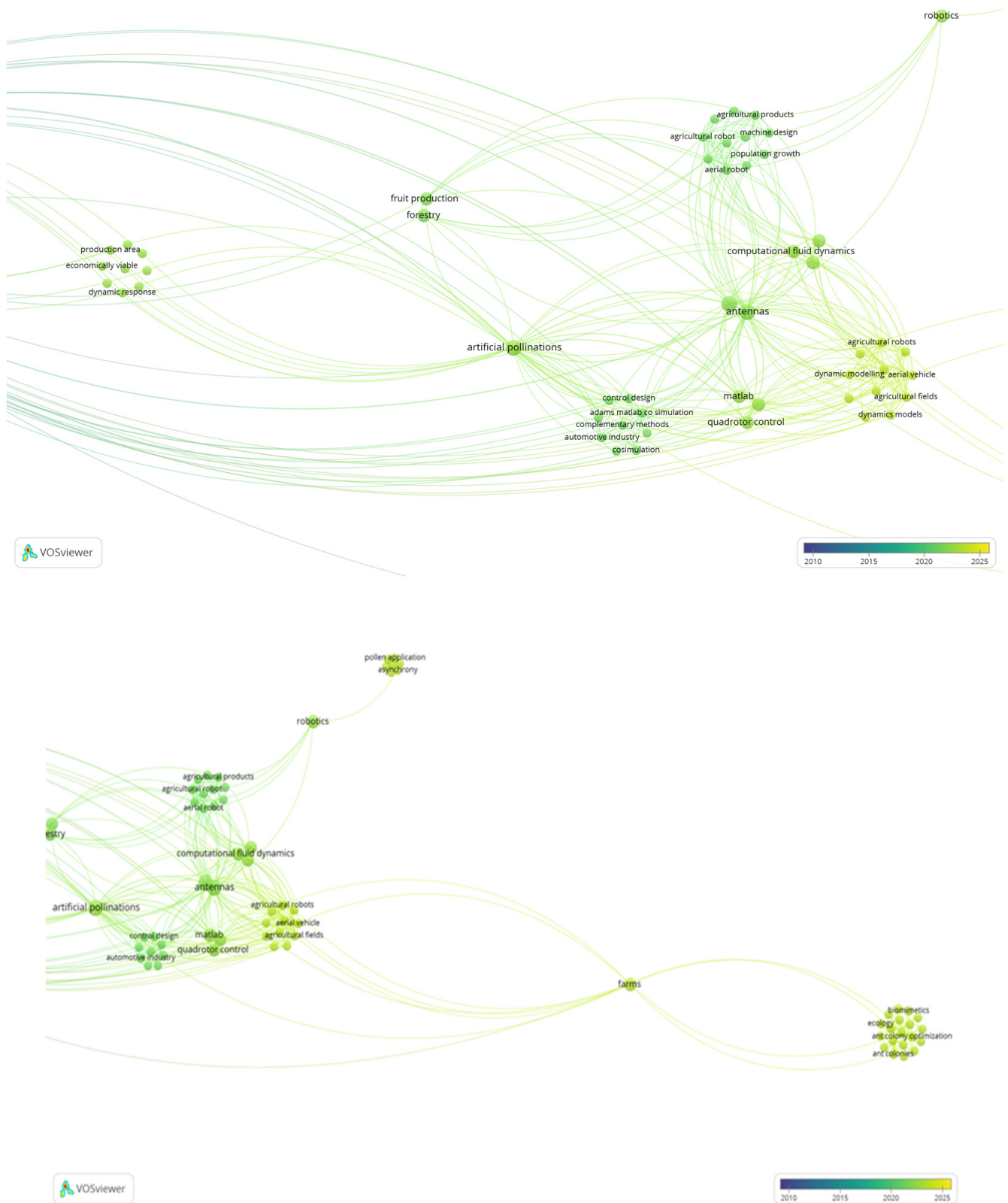


Figure 3. Co-keyword network visualization based on “Artificial” and “Pollination and Technologies and Walnut”. Results are based on Cluster 1, which is defined by 52 keywords (items).

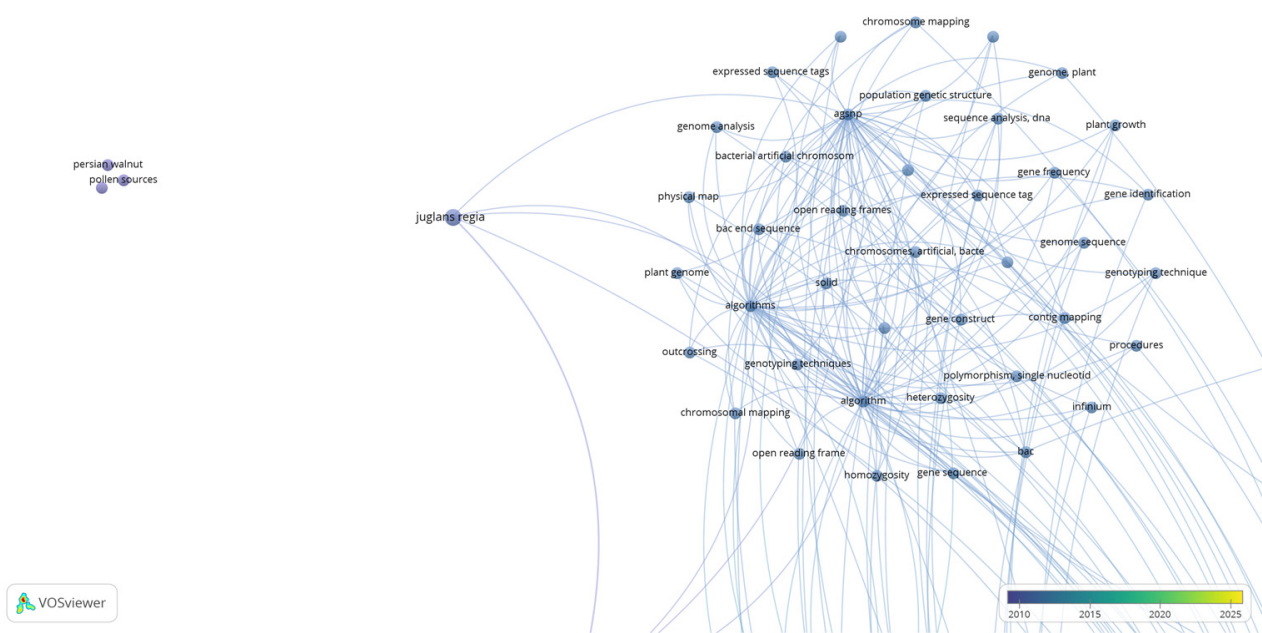


Figure 4. Co-keyword network visualization based on “Artificial” and “Pollination and Technologies and Walnut”. Results are based on Cluster 2, which is defined by 45 keywords (items).

Based on simulation methodologies for controlling a quadrotor, MATLAB provides a solution for the standard flower pollination algorithm, with more information on nature optimization algorithms provided in a book entitled *Nature-Inspired Optimization Algorithms*. The flower pollination algorithm (FPA) is a type of optimization algorithm based on the behavior of flowering plants [47]. It is a population-based metaheuristic algorithm that has been modified and hybridized to perform efficiently across a range of optimization problems. One such modification involves the integration of the bee pollinator concept, which has shown promising results in solving the data clustering problem [48]. A modified version of the FPA, which utilizes the crossover technique for resolving multidimensional knapsack problems, was also developed [49]. Keeping the abovementioned facts in view, we believe that an algorithm such as the FPA should be used for a pollination robot to prevent or reduce the risk of walnut blight disease.

In addition to the FPA, the bacterial microbiota associated with flower pollen is influenced by the pollen type and exhibits a significant degree of diversity and species specificity [49]. So, for microbe–pollen interactions, presumably we must know the sequencing of 16S rRNA gene amplicon libraries to identify dominant microbial phyla and the core microbiome of pollen from different walnut cultivars. Since pollen-associated bacteria may have a potential impact on walnut blight disease, pollen microbial communities from different walnut cultivars need to be identified. This information could play a significant role in the flower pollination algorithm and development of a strategy that leads to new valuable information on walnut cultivar microbe–pollen interactions during pollination.

Moreover, considering the algorithm-based approach, it is necessary to provide criteria for the aerial robot path. Basic simulation steps helped to determine pollen streams under a quadrotor unmanned aerial vehicle [35]. These steps were the following:

1. Design and fabrication of quadrotor components For the quadrotor to be able to carry and distribute pollen over trees, the quadrotor components needed to be selected correctly, i.e., the body size was chosen based on the pollen tank. The objective of the quadrotor was to pollinate, so the tank was equipped with a nozzle with four holes to allow pollen to fall uniformly. It was recommended that the propulsion system use a motor with a lower current. The electronic device contained various sensors, such as a compass, a global positioning system (GPS), a barometer, and connections for the flight control system [35].

2. Modeling and Control Computational fluid dynamic (CFD) software was used to simulate the airflow beneath the UAV. This simulation assisted in identifying the pollen streams under the robot so that the released pollen could be directed toward the target trees. The quadrotor controller was designed and simulated in MATLAB to ensure system stability [35].
3. Pollination of walnut trees Different mixtures of pollen diluted with talcum powder were distributed over various groups of trees using the quadrotor. The results achieved on the trees were measured after a few months to evaluate the benefits of the proposed system [35].

In the context of pollination supply modeling, MATLAB is utilized to predict pollination services like pollinator visitation rates, fruit set, and pollen deposition [50]. These predictions can help to optimize pollination services in the field and support the development of effective agri-environmental schemes or conservation initiatives. Bayesian models, particularly in MATLAB, are pivotal in pollination research for various applications [51]. These models aid in reconstructing plant–pollinator networks from observational data by separating structure from noise, estimating the expected number of visits between specific plant–pollinator species pairs, and predicting pollinator abundance under different scenarios. In summary, research shows that the integration of Bayesian models with MATLAB provides a powerful tool for researchers to analyze plant–pollinator interactions, predict pollinator abundance, optimize pollination services, and make informed decisions regarding conservation strategies and agricultural practices based on reliable data-driven insights [50,51].

5. Machine Learning and Pollination

Machine learning (ML) can play a significant role in the context of pollination, offering innovative solutions and insight to address challenges and optimize pollination processes. Below are several ways in which machine learning is applied to pollination:

1. Predictive modeling: ML algorithms can analyze historical data on pollination success, environmental conditions, crop yields, and phenology to create predictive models. These models can help to predict optimal times for pollination, considering factors such as weather patterns, bloom asynchrony, and the availability of pollinators [52–54].
2. Automated monitoring: ML-powered monitoring systems can analyze images or sensor data to track the health and development of crops. This real-time monitoring allows for the early detection of issues related to pollination, such as low pollination rates or the presence of pests that could affect pollinators [55,56].
3. Optimizing pollination strategies: Machine learning algorithms can optimize artificial pollination strategies based on a variety of factors [57,58]. These include the type of crop, environmental conditions, and the efficiency of different pollination methods [59,60]. This can lead to more targeted and effective pollination efforts [61].
4. Identification of pollinator behavior: ML can be used to analyze the behavior of natural pollinators, such as bees, by processing video footage or sensor data. Understanding pollinator behavior can provide insight into their preferences, movement patterns, and efficiency in pollinating specific crops [62,63].
5. Genetic analysis: Machine learning techniques can analyze genetic data related to plant characteristics, including traits associated with pollination [64,65]. This information can contribute to breeding programs aimed at developing crops that are more resilient to environmental challenges and more compatible with artificial pollination methods [66,67].
6. Decision support systems: ML-based decision support systems can assist farmers in making informed choices related to pollination strategies. These systems can consider a range of variables and provide recommendations for optimal pollination practices [68,69].

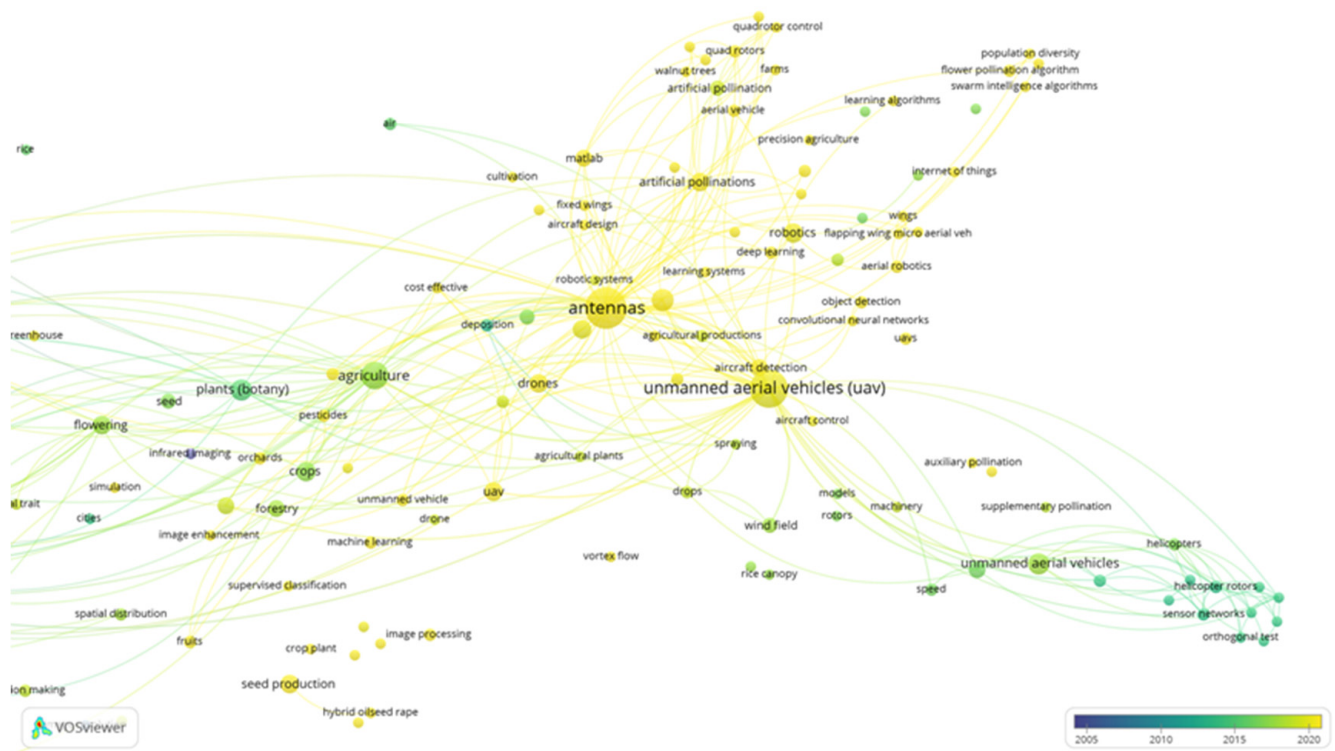


Figure 6. Co-keyword network visualization based on “Aerial” and “Pollination”.

6. Walnut Pollination: Model to Prevent or Reduce the Risk of the Walnut Blight Disease (Our View)

Developing a walnut pollination model involves creating a predictive framework that considers various factors influencing the pollination of walnut trees [36,48]. Keeping in mind the success of the walnut pollination model (Figures 1–6) and based on the above paragraphs, we illustrate the following flowchart (6 steps, state of the art flowchart), which provides a general guideline for conducting a state-of-the-art analysis for a pollination robot to prevent or reduce the risk of walnut blight disease. The development of autonomous visual navigation for a flower pollination drone has been presented in a paper published in the journal *Machines* [73]. This technology could enable drones to autonomously approach flowers and perform pollination tasks.

State of the Art Flowchart

1st Step. Walnuts: budbreak, bloom, and pollination growth stages

- 1.1. Identify spring bud break, leaf emergence, and anthesis growth stages of the walnut cultivars.
- 1.2. Understand how plants grow and develop during bud break, anthesis, and pollination.
- 1.3. Understand protandrous and protogynous mechanisms prior to pollen shedding.

2nd Step. Study the bacterial microbiota associated with flower pollen

- 2.1. Identify the “core” pollen microbiota.
- 2.2. Compare bacterial abundance and diversity between walnut cultivars.
- 2.3. Assess the impact of the pollination type on the variability of the flower pollen microbiota.
- 2.4. Estimate the role of *X. arboricola* pv. *juglandis* in stigma exposed to contaminated pollen.

3rd Step. State of Walnut Pollination – Develop Pollination Algorithms

- 3.1. Check reservoir cultivars, i.e., Chandler, for the presence of inoculum.

- 3.2. Check for conditions that encourage the disease to spread, such as moisture, and especially the combined action of wind and rain.
- 3.3. Check for air-borne inoculum when catkins open.
- 3.4. Check for pistil-late flowers.
- 3.5. Collect and store uninfected pollen.

4th Step. State of Walnut Pollination – Develop Pollination Algorithms

- 4.1. Set genetic algorithm for ‘mutation’, ‘selection’, and ‘recombination’ based on pollen–microbe interaction.
- 4.2. Use metaheuristic algorithms, which include differential evolution.
- 4.3. Use the appropriate flower pollination algorithm that is suitable and inspired by the process of pollination.

5th Step. Design of an Autonomous Precision Pollination Robot

- 5.1. Quadrotor (pollinating drone) to carry pollen grains.
- 5.2. Pollinating drone to make an ideal delivery system, landing on the pistil of a flower to result in pollination.
- 5.3. Drone technique would need some refinement in localization, mapping, and control.
- 5.4. Metaheuristic optimization algorithm to find the best (feasible) solution out of all possible solutions to the pollination optimization problem.

6th Step. Algorithm Optimization

- 6.1. Analyze historical data on pollination success, environmental conditions, and crop yields (machine learning).
- 6.2. Tune flower pollination algorithm parameters (formulating the above steps, mainly steps 1 and 2, due to climate change).
- 6.3. Start the process from step 3 and improve the equipment at steps 4 and 5.

7. Conclusions

Evidence to date is convincing that microbes such as the bacterium *X. arboricola* pv. *juglandis* occur in pollen and influence walnut blight disease. Research shows that *X. arboricola* pv. *juglandis*, the causative agent of walnut blight, can affect the leaves, stems, and nuts of the tree, and even the pollen. In this review, we present a flow diagram of a new artificial pollinator that we assume can reduce the spread of the disease in orchards. As we have seen in this study, we can conclude that the walnut pollination model and walnut blight disease are complex problems that require solutions due to the interaction of several factors. Significant progress in understanding these dynamic processes is reported in the literature utilizing various scientific approaches. Considering the knowledge needed for a solution to overcome this bacterial plant infection, the VOS viewer results suggest that the foundation for a new, more sophisticated and efficient walnut pollination model to prevent or reduce the risk of walnut blight disease is possible. So, the purpose of the ideas presented here was to facilitate a description of such a pollination model with fundamental components, including the identification of the ‘core’ pollen microbiota, specifying an appropriate flower pollination algorithm, design of an autonomous precision pollination robot, and minimizing the average errors of flower pollination algorithm parameters through machine learning and meta-heuristic algorithms.

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References

- Polito, V.S.; Pinney, K.; Weinbaum, S.; Aradhya, M.K.; Dangl, J.; Yanknin, Y.; Grant, J.A. Walnut pollination dynamics: Pollen flow in walnut orchards. *Acta Hort.* **2005**, *705*, 465–472. [\[CrossRef\]](#)
- Ark, P.A. Further evidence of pollen dissemination of walnut blight. *Phytopathology* **1944**, *34*, 329–334.
- Kałużna, M.; Fischer-Le Saux, M.; Pothier, J.F.; Jacques, M.A.; Obradović, A.; Tavares, F.; Stefani, E. *Xanthomonas arboricola* pv. *juglandis* and pv. *corylina*: Brothers or distant relatives? Genetic clues, epidemiology, and insights for disease management. *Mol. Plant Pathol.* **2021**, *22*, 1481–1499. [\[CrossRef\]](#)
- Mulrean, E.N.; Schroth, M.N. Ecology of *Xanthomonas campestris* pv. *juglandis* on Persian (English) Walnuts. *Phytopathology* **1982**, *72*, 434–438. [\[CrossRef\]](#)
- Lang, M.D.; Evans, K. Epidemiology and status of walnut blight in Australia. *J. Plant Pathol.* **2010**, *92*, 49–56.
- Giovanardi, D.; Bonneau, S.; Gironde, S.; Saux, M.F.; Manceau, C.; Stefani, E. Morphological and genotypic features of *Xanthomonas arboricola* pv. *juglandis* populations from walnut groves in Romagna region, Italy. *Eur. J. Plant Pathol.* **2016**, *145*, 1–16. [\[CrossRef\]](#)
- Tontou, R.; Giovanardi, D.; Stefani, E. Pollen as a possible pathway for the dissemination of *Pseudomonas syringae* pv. *actinide* and bacterial canker of kiwifruit. *Phytopathol. Mediterr.* **2014**, *53*, 333–339. [\[CrossRef\]](#)
- Van der Zwet, T.; Bell, R.L. Survival of *Erwinia amylovora* on apple and pear pollen. *Acta Hort.* **1992**, *338*, 111–112. [\[CrossRef\]](#)
- Garcin, A.; El-Maataoui, M.; Tichadou, S.; Prunet, J.P.; Ginibre, T.; Penet, C. Walnut blight, new knowledge for an old disease: Summary of research (1995–2000). *Infos-Ctifl* **2001**, *171*, 27–30.
- Giovanardi, D.; Dallai, D.; Stefani, E. Population features of *Xanthomonas arboricola* pv. *juglandis* and epidemiology of walnut blight in Romagna (Italy). In Proceedings of the Petria 13th Congress of the Mediterranean Phytopathological Union, Rome, Italy, 20–25 June 2010; Volume 20, pp. 96–97.
- Matsumoto, D.; Shimizu, S.; Shimazaki, A.; Ito, K.; Taira, S. Effects of Self-pollen Contamination in Artificial Pollination on Fruit Set of ‘Fuji Murasaki’ Akebia trifoliata. *Hortic. J.* **2022**, *91*, 431–436. [\[CrossRef\]](#)
- Everett, K.; Cohen, D.; Pushparajah, I.; Vergara, M.; Curtis, C.; Larsen, N.; Jia, Y. Heat treatments to kill *Pseudomonas syringae* pv. *actinidiae* on contaminated pollen. *N. Z. Plant Prot.* **2012**, *65*, 8–18. [\[CrossRef\]](#)
- Dingley, A.; Anwar, S.; Kristiansen, P.; Warwick, N.W.; Wang, C.; Sindel, B.M.; Cazzonelli, C.I. Precision Pollination Strategies for Advancing Horticultural Tomato Crop Production. *Agronomy* **2022**, *12*, 518. [\[CrossRef\]](#)
- Hiraguri, T.; Kimura, T.; Endo, K.; Ohya, T.; Takanashi, T.; Shimizu, H. Shape classification technology of pollinated tomato flowers for robotic implementation. *Sci. Rep.* **2023**, *13*, 2159. [\[CrossRef\]](#) [\[PubMed\]](#)
- Sánchez-Molina, J.; Rodríguez, F.; Moreno, J.; Sánchez-Hermosilla, J.; Giménez, A. Robotics in greenhouses. Scoping review. *Comput. Electron. Agric.* **2024**, *219*, 108750. [\[CrossRef\]](#)
- Gudowska, A.; Cwajna, A.; Marjańska, E.; Moroń, D. Pollinators enhance the production of a superior strawberry—A global review and meta-analysis. *Agric. Ecosyst. Environ.* **2024**, *362*, 108815. [\[CrossRef\]](#)
- Rice, C.R.; McDonald, S.T.; Shi, Y.; Gan, H.; Lee, W.S.; Chen, Y.; Wang, Z. Perception, Path Planning, and Flight Control for a Drone-Enabled Autonomous Pollination System. *Robotics* **2022**, *11*, 144. [\[CrossRef\]](#)
- Yamada, N.; Hiraguri, T.; Shimizu, H.; Kimura, T.; Shimada, T.; Shibasaki, A.; Takemura, Y. Drone Flight Experiment using RTK Positioning for Pear Pollination. In Proceedings of the 2023 International Conference on Consumer Electronics—Taiwan (ICCE-Taiwan), PingTung, Taiwan, 17–19 July 2023; pp. 655–656. [\[CrossRef\]](#)
- Chevallier, A.; Bray, O.; Prunet, J.P.; Giraud, M. Factors influencing walnut blight symptoms emergence and development. *Acta Hort.* **2010**, *861*, 473–478. [\[CrossRef\]](#)
- Vagelas, I.; Rumbos, C.I.; Tsiantos, J.A. Variation in disease development among persian walnut cultivars, selections and crosses when inoculated with *Xanthomonas arboricola* pv. *juglandis* in Greece. *J. Plant Pathol.* **2012**, *94*, 57–61. [\[CrossRef\]](#)
- Moragrega, C.; Llorente, I. Effects of leaf wetness duration, temperature, and host phenological stage on infection of walnut by *Xanthomonas arboricola* pv. *juglandis*. *Plants* **2023**, *12*, 2800. [\[CrossRef\]](#)
- Lindow, S.; Olson, W.; Buchner, R. Colonization of dormant walnut buds by *Xanthomonas arboricola* pv. *juglandis* is predictive of subsequent disease. *Phytopathology* **2014**, *104*, 1163–1174. [\[CrossRef\]](#)
- Buchner, R.P.; Gilles, C.; Olson, W.H.; Adaskaveg, J.E.; Lindow, S.E.; Koutsoukis, R. Spray timing and materials for walnut blight (*Xanthomonas campestris* pv. *juglandis*, *Xanthomonas arboricola* pv. *juglandis*) control in northern California USA. *Acta Hort.* **2010**, *861*, 457–464. [\[CrossRef\]](#)
- Adaskaveg, J.E.; Förster, H.; Thompson, D.; Enns, J.; Connell, J.; Buchner, R. Epidemiology and management of walnut blight. *Walnut Res. Rep.* **2009**, 241–257.
- Miller, P.W.; Bollen, W.B. *Walnut Bacteriosis and Its Control*; Agricultural Experiment, Station Technical Bulletin 9; United States Department of Agriculture Bureau of Plant Industry, Soils and Agricultural Engineering, Oregon State College: Corvallis, OR, USA, 1946.

26. Franchi, G.G.; Piotto, B.; Nepi, M.; Baskin, C.C.; Baskin, J.M.; Pacini, E. Pollen and seed desiccation tolerance in relation to degree of developmental arrest, dispersal, and survival. *J. Exp. Bot.* **2011**, *62*, 5267–5281. [[CrossRef](#)] [[PubMed](#)]
27. Donati, I.; Cellini, A.; Sangiorgio, D.; Vanneste, J.; Scortichini, M.; Balestra, G.; Spinelli, F. *Pseudomonas syringae* pv. *actinidiae*: Ecology, infection dynamics and disease epidemiology. *Microb. Ecol.* **2020**, *80*, 81–102. [[CrossRef](#)] [[PubMed](#)]
28. Farina, W.M.; Arenas, A.; Díaz, P.C.; Susic Martin, C.; Corriale, M.J. In-hive learning of specific mimic odours as a tool to enhance honey bee foraging and pollination activities in pear and apple crops. *Sci. Rep.* **2022**, *12*, 20510. [[CrossRef](#)] [[PubMed](#)]
29. Pedroncelli, A.; Puopolo, G. This tree is on fire: A review on the ecology of *Erwinia amylovora*, the causal agent of fire blight disease. *J. Plant Pathol.* **2023**. [[CrossRef](#)]
30. Broussard, M.A.; Coates, M.; Martinsen, P. Artificial pollination technologies: A review. *Agronomy* **2023**, *13*, 1351. [[CrossRef](#)]
31. Castro, H.; Siopa, C.; Casais, V.; Castro, M.; Loureiro, J.; Gaspar, H.; Castro, S. Pollination as a key management tool in crop production: Kiwifruit orchards as a study case. *Sci. Hort.* **2021**, *290*, 110533. [[CrossRef](#)]
32. Wurz, A.; Grass, I.; Tschardt, T. Hand pollination of global crops—A systematic review. *Basic Appl. Ecol.* **2021**, *56*, 299–321. [[CrossRef](#)]
33. Shimizu, H. Advanced technologies for pollination in plant factories. In *Plant Factory Using Artificial Light*; Elsevier: Amsterdam, The Netherlands, 2019; pp. 185–192. [[CrossRef](#)]
34. Frachon, L.; Stirling, S.; Schiestl, F.P.; Dudareva, N. Combining biotechnology and evolution for understanding the mechanisms of pollinator attraction. *Curr. Opin. Biotechnol.* **2021**, *70*, 213–219. [[CrossRef](#)]
35. Mazinani, M.; Zarafshan, P.; Dehghani, M.; Vahdati, K.; Etezadi, H. Design and analysis of an aerial pollination system for walnut trees. *Biosyst. Eng.* **2023**, *225*, 83–98. [[CrossRef](#)]
36. Potts, S.G.; Neumann, P.; Vaissière, B.E.; Vereecken, N.J. Robotic bees for crop pollination: Why drones cannot replace biodiversity. *Sci. Total Environ.* **2018**, *642*, 665–667. [[CrossRef](#)]
37. Chen, P.; Douzals, J.; Lan, Y.; Cotteux, E.; Delpuech, X.; Pouxviel, G.; Zhan, Y. Characteristics of unmanned aerial spraying systems and related spray drift: A review. *Front. Plant Sci.* **2022**, *13*, 870956. [[CrossRef](#)]
38. Zhang, Y.; Huang, X.; Lan, Y.; Wang, L.; Lu, X.; Yan, K.; Deng, J.; Zeng, W. Development and prospect of UAV-based aerial electrostatic spray technology in China. *Appl. Sci.* **2021**, *11*, 4071. [[CrossRef](#)]
39. Ge, C.; Dunno, K.D.; Singh, M.; Yuan, L.; Lu, L. Development of a drone’s vibration, shock, and atmospheric profiles. *Appl. Sci.* **2021**, *11*, 5176. [[CrossRef](#)]
40. Russell, A.L.; Papaj, D.R. Artificial pollen dispensing flowers and feeders for bee behaviour experiments. *J. Pollinat. Ecol.* **2016**, *18*, 13–22. [[CrossRef](#)]
41. Li, K.; Huo, Y.; Liu, Y.; Shi, Y.; He, Z.; Cui, Y. Design of a lightweight robotic arm for kiwifruit pollination. *Comput. Electron. Agric.* **2022**, *198*, 107114. [[CrossRef](#)]
42. Eck, N.J.; Waltman, L. Software survey: VOSviewer, a computer program for bibliometric mapping. *Scientometrics* **2009**, *84*, 523–538. [[CrossRef](#)]
43. Orduña-Malea, E.; Costas, R. Link-based approach to study scientific software usage: The case of VOSviewer. *Scientometrics* **2021**, *126*, 8153–8186. [[CrossRef](#)]
44. Arruda, H.; Silva, É.R.; Lessa, M.; Proença, D.; Bartholo, R. VOSviewer and Bibliometrix. *J. Med. Libr. Assoc.* **2022**, *110*, 392–395. [[CrossRef](#)] [[PubMed](#)]
45. Vagelas, I.; Leontopoulos, S. A bibliometric analysis and a citation mapping process for the role of soil recycled organic matter and microbe interaction due to climate change using scopus database. *Agric. Eng.* **2023**, *5*, 581–610. [[CrossRef](#)]
46. Lykas, C.; Vagelas, I. Innovations in agriculture for sustainable Agro-systems. *Agronomy* **2023**, *13*, 2309. [[CrossRef](#)]
47. Mergos, P.E.; Yang, X. Flower pollination algorithm parameters tuning. *Soft Comput.* **2021**, *25*, 14429–14447. [[CrossRef](#)] [[PubMed](#)]
48. Wang, R.; Zhou, Y.; Qiao, S.; Huang, K. Flower pollination algorithm with bee pollinator for cluster analysis. *Inf. Proc. Lett.* **2016**, *116*, 1–14. [[CrossRef](#)]
49. Ambika Manirajan, B.; Ratering, S.; Rusch, V.; Schwiertz, A.; Geissler-Plaum, R.; Cardinale, M.; Schnell, S. Bacterial microbiota associated with flower pollen is influenced by pollination type, and shows a high degree of diversity and species-specificity. *Environ. Microbiol.* **2016**, *18*, 5161–5174. [[CrossRef](#)] [[PubMed](#)]
50. Akwasi, A.M.; Wei, X.; Duku, O.A. Observer controller-based structure for a modified flower pollination algorithm for wind power generation. *Int. J. Autom. Control* **2024**, *18*, 53–86. [[CrossRef](#)]
51. Hemalatha, S.; Johny Renoald, A.; Banu, G.; Indirajith, K. Design and investigation of PV string/central architecture for bayesian fusion technique using grey wolf optimization and flower pollination optimized algorithm. *Energy Convers. Manag.* **2023**, *286*, 117078. [[CrossRef](#)]
52. Campbell, D.R. Predicting plant reproductive success from models of competition for pollination. *Oikos* **1986**, *47*, 257–266. [[CrossRef](#)]
53. Sáez, A.; di Virgilio, A.; Tiribelli, F.; Geslin, B. Simulation models to predict pollination success in apple orchards: A useful tool to test management practices. *Apidologie* **2018**, *49*, 551–561. [[CrossRef](#)]
54. Stock, M.; Piot, N.; Vanbesien, S.; Meys, J.; Smagghe, G.; Baets, B.D. Pairwise learning for predicting pollination interactions based on traits and phylogeny. *Ecol. Model.* **2021**, *451*, 109508. [[CrossRef](#)]
55. Pegoraro, L.; Hidalgo, O.; Leitch, I.J.; Pellicer, J.; Barlow, S.E. Automated video monitoring of insect pollinators in the field. *Emerg. Top Life Sci.* **2020**, *4*, 87–97. [[CrossRef](#)] [[PubMed](#)]

56. Ngo, T.N.; Rustia, D.J.; Yang, E.; Lin, T. Automated monitoring and analyses of honey bee pollen foraging behavior using a deep learning-based imaging system. *Comput. Electron. Agric.* **2021**, *187*, 106239. [[CrossRef](#)]
57. DeVetter, L.W.; Chabert, S.; Milbrath, M.O.; Mallinger, R.E.; Walters, J.; Isaacs, R.; Galinato, S.P.; Kogan, C.J.; Brouwer, K.; Melathopoulos, A.; et al. Toward evidence-based decision support systems to optimize pollination and yields in highbush blueberry. *Front. Sustain. Food Syst.* **2022**, *6*, 1006201. [[CrossRef](#)]
58. Menzel, C.M. Fruit set is moderately dependent on insect pollinators in strawberry and is limited by the availability of pollen under natural open conditions. *J. Hortic. Sci. Biotechnol.* **2023**, *98*, 685–714. [[CrossRef](#)]
59. Yang, M.; Lyu, H.; Zhao, Y.; Sun, Y.; Pan, H.; Sun, Q.; Chen, J.; Qiang, B.; Yang, H. Delivery of pollen to forsythia flower pistils autonomously and precisely using a robot arm. *Comput. Electron. Agric.* **2023**, *214*, 108274. [[CrossRef](#)]
60. Hiraguri, T.; Shimizu, H.; Kimura, T.; Matsuda, T.; Maruta, K.; Takemura, Y.; Ohya, T.; Takanashi, T. Autonomous drone-based pollination system using AI classifier to replace bees for greenhouse tomato cultivation. *IEEE Access* **2023**, *11*, 99352–99364. [[CrossRef](#)]
61. Cong, W.; Dupont, Y.L.; Søgaard, K.; Eriksen, J. Optimizing yield and flower resources for pollinators in intensively managed multi-species grasslands. *Agric. Ecosyst. Environ.* **2020**, *302*, 107062. [[CrossRef](#)]
62. Wajnberg, E.; Tel-Zur, N.; Shapira, I.; Leber, Y.; Lev-Yadun, S.; Zurgil, U.; Reisman-Berman, O.; Keasar, T. Pollinator behavior drives sexual specializations in the hermaphrodite flowers of a heterodichogamous tree. *Front. Plant Sci.* **2019**, *10*, 1315. [[CrossRef](#)]
63. Knauer, A.C.; Kokko, H.; Schiestl, F.P. Pollinator behaviour and resource limitation maintain honest floral signalling. *Funct. Ecol.* **2021**, *35*, 2536–2549. [[CrossRef](#)]
64. Yuan, Y.; Byers, K.J.; Bradshaw, H.D. The genetic control of flower-pollinator specificity. *Curr. Opin. Plant Biol.* **2013**, *16*, 422–428. [[CrossRef](#)]
65. Rose, J.P.; Sytsma, K.J. Complex interactions underlie the correlated evolution of floral traits and their association with pollinators in a clade with diverse pollination systems. *Evolution* **2021**, *75*, 1431–1449. [[CrossRef](#)]
66. Feigs, J.T.; Holzhauser, S.I.; Huang, S.; Brunet, J.; Diekmann, M.; Hedwall, P.; Kramp, K.; Naaf, T. Pollinator movement activity influences genetic diversity and differentiation of spatially isolated populations of clonal forest herbs. *Front. Ecol. Evol.* **2022**, *10*, 908258. [[CrossRef](#)]
67. Opedal, Ø.H.; Pérez-Barrales, R.; Brito, V.L.; Muchhala, N.; Capó, M.; Dellinger, A.S. Pollen as the link between floral phenotype and fitness. *Am. J. Bot.* **2023**, *110*, e16200. [[CrossRef](#)]
68. Barons, M.J.; Shenvi, A. Where the bee sucks—A dynamic bayesian network approach to decision support for pollinator abundance strategies. *arXiv* **2022**, arXiv:2212.03179. [[CrossRef](#)]
69. Lonsdorf, E.V.; Kremen, C.; Ricketts, T.H.; Winfree, R.; Williams, N.M.; Greenleaf, S.S. Modelling pollination services across agricultural landscapes. *Ann. Bot.* **2009**, *103*, 1589–1600. [[CrossRef](#)] [[PubMed](#)]
70. Drucker, D.P.; Salim, J.A.; Trekels, M.; Groom, Q.J.; Parr, C.S.; Soares, F.M.; Agostini, K.; Saraiva, A.; Molloy, L.; Hodson, S.; et al. Plant-pollinator interaction data: A case study of the WorldFAIR project. *Biodivers. Inf. Sci. Stand.* **2022**, *6*, e94310. [[CrossRef](#)]
71. Salim, J.A.; Zermoglio, P.F.; Drucker, D.P.; Soares, F.M.; Saraiva, A.M.; Agostini, K.; Freitas, L.; Wolowski, M.; Rech, A.R.; Maués, M.M.; et al. Plant-pollinator vocabulary—A contribution to interaction data standardization. *Biodivers. Inf. Sci. Stand.* **2021**, *5*, e75636. [[CrossRef](#)]
72. Salim, J.A.; Saraiva, A.M.; Zermoglio, P.F.; Agostini, K.; Wolowski, M.; Drucker, D.P.; Soares, F.M.; Bergamo, P.J.; Varassin, I.G.; Freitas, L.; et al. Data standardization of plant–pollinator interactions. *GigaScience* **2022**, *11*, giac043. [[CrossRef](#)] [[PubMed](#)]
73. Hulens, D.; Van Ranst, W.; Cao, Y.; Goedemé, T. Autonomous Visual Navigation for a Flower Pollination Drone. *Machines* **2022**, *10*, 364. [[CrossRef](#)]

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